

Prepared in cooperation with the Great Lakes Restoration Initiative

# Occurrence and Distribution of Fecal Indicator Bacteria and Gene Markers of Pathogenic Bacteria in Great Lakes Tributaries, March–October 2011



Open-File Report 2015–1013



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By Angela K. Brennan, Heather E. Johnson, Alex R. Totten, and Joseph W. Duris

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**U.S. Department of the Interior**  
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# Contents

Abstract.....	1
Introduction.....	1
Purpose and Scope .....	2
Description of Study Area .....	2
Sampling and Analytical Methods .....	2
Site Selection.....	2
Sample Collection .....	2
Microbiological Analyses .....	6
Bacteria Enumeration, Enrichment, and Preservation.....	6
DNA Extraction.....	6
Polymerase Chain Reaction Assays for Pathogen Detection .....	6
Results of Microbiological Analyses .....	6
Occurrence and Distribution of Fecal Indicator Bacteria .....	6
Occurrence and Distribution of Bacterial Pathogen Gene Markers .....	8
Quality-Control Results .....	10
Summary.....	10
Acknowledgements.....	11
References Cited.....	11
Appendixes .....	13

## Figures

1. Graph showing land cover surrounding tributary sites sampled as part of the Great Lakes Restoration Initiative (GLRI) pathogens study.....	3
2. Map showing Great Lakes Restoration Initiative (GLRI) USGS tributary sampling locations and number of samples collected for pathogens study, 2011 .....	4
3. Graph showing frequency of samples testing positive for pathogen gene markers .....	8
4. Graph showing percentage of pathogen gene markers in relation to water quality criteria.....	9

## Tables

1. Great Lakes Restoration Initiative (GLRI) pathogens study sampling locations, 2011 .....	5
2. Growth media, target organisms, and incubation temperature requirements for the Great Lakes Restoration Initiative (GLRI) pathogens study.....	7
3. Selected pathogen gene targets and virulence traits of the organism.....	7
4. Summary of the frequency of pathogen gene detections in a single sample.....	8
5. Percentage of samples that tested positive for pathogenic gene markers during high-flow and normal-flow conditions .....	9
6. Relative standard deviation of field replicates for quantitative microbial methods in the Great Lakes Restoration Initiative (GLRI) pathogens study.....	10
7. Frequency of agreement between field replicates for qualitative microbial methods in the Great Lakes Restoration Initiative (GLRI) pathogens study .....	10

## Conversion Factors

<b>Multiply</b>	<b>By</b>	<b>To obtain</b>
Length		
mile (mi)	1.609	kilometer (km)
Area		
square mile (mi <sup>2</sup> )	2.590	square kilometer (km <sup>2</sup> )
Volume		
milliliter (mL)	1,000	microliter (μL)

Temperature in degrees Celsius (°C) may be converted to degrees Fahrenheit (°F) as follows:  $^{\circ}\text{F}=(1.8\times^{\circ}\text{C})+32$

Horizontal coordinate information is referenced to North American Datum of 1983 (NAD 83).

Concentrations of fecal indicator bacteria are given in colony-forming units per 100 milliliters (CFU/100 mL).

## Abbreviations used in report

CDC	Centers for Disease Control and Prevention
CFU	colony-forming unit
DNA	Deoxyribonucleic acid
<i>E. coli</i>	<i>Escherichia coli</i>
EWI	equal-width-increment
FIB	Fecal indicator bacteria
GLRI	Great Lakes Restoration Initiative
MI-BaRL	USGS Michigan Bacteriological Research Laboratory
MI-WSC	USGS Michigan Water Science Center
PBS	phosphate buffered saline
PCR	polymerase chain reaction
QC	quality control
RSD	relative standard deviation
STEC	Shiga-toxin producing <i>Escherichia coli</i>
EPA	U.S. Environmental Protection Agency
USGS	U.S. Geological Survey
WQC	water quality criteria



# Occurrence and Distribution of Fecal Indicator Bacteria and Gene Markers of Pathogenic Bacteria in Great Lakes Tributaries, March–October 2011

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## Abstract

From March through October 2011, the U.S. Geological Survey (USGS), conducted a study to determine the frequency of occurrence of pathogen gene markers and densities of fecal indicator bacteria (FIB) in 22 tributaries to the Great Lakes. This project was funded as part of the Great Lakes Restoration Initiative (GLRI) and included sampling at 22 locations throughout 6 states that border the Great Lakes.

A total of 177 environmental samples were collected at USGS streamgaging stations during both normal-flow and high-flow conditions and were analyzed by the Michigan Bacteriological Research Laboratory at the USGS Water Science Center in Lansing, Michigan.

Water samples were analyzed for the presence of FIB concentrations (FIB; fecal coliform bacteria, *Escherichia coli* [*E. coli*], and enterococci) by using membrane filtration and serial dilution methods. The resulting enrichments from standard culturing of the samples were then analyzed by using polymerase chain reaction (PCR) to determine the occurrence of pathogen gene markers for *Shigella* species, *Campylobacter jejuni* and *coli*, *Salmonella* species, and pathogenic *E. coli*, including Shiga toxin-producing *E. coli* (STEC).

## Introduction

In 2009, the Great Lakes Restoration Initiative (GLRI) was started as an interagency initiative to protect and restore the Great Lakes. This report describes data collected in 2011 as part of the U.S. Geological Survey (USGS) GLRI effort led by the Michigan Water Science Center (MI-WSC), in conjunction with USGS Water Science Centers in Indiana, Minnesota, Ohio, New York, and Wisconsin, and complements the ongoing USGS GLRI Great Lakes Nutrient and Sediment Loadings study (GLRI Loadings study) being conducted at the same sampling locations.

The microbiological monitoring network for this study consisted of 22 sampling locations, representing multiple land-use types, drainage areas, and soil characteristics typical of the Great Lakes region. Samples were collected to better understand the occurrence and distribution of fecal indicator bacteria (FIB) and selected bacterial pathogens that are commonly associated with waterborne illnesses.

Within the broad FIB groups (*Escherichia coli* [*E. coli*], fecal coliforms, and enterococci), there are specific strains of bacteria that are pathogenic (infection causing) to humans. The bacterial pathogen gene markers evaluated in this study included genes from the genera *Shigella*, *Campylobacter jejuni* and *coli* and *Salmonella*. In addition, several genes were also evaluated from a class of pathogenic *E. coli* known as Shiga toxin-producing *E. coli* (STEC), which includes *E. coli* O157:H7.

Fecally derived bacterial pathogens have long been known to be a health threat to humans, and exposure to these pathogens can result in illness and occasionally even death (Centers for Disease Control and Prevention [CDC], 2012b). STEC, including *E. coli* O157:H7, can cause illness ranging from mild intestinal disease to severe kidney complications and death in animals and humans (CDC, 2012d). *Shigella*'s pathogenicity functions similarly to that of STEC; however, *Shigella* mainly affects humans. *Campylobacter* is one of the most common causes of diarrheal illness in the United States, and symptoms include cramping, abdominal pain, and fever (CDC, 2012a). *Salmonella* infection can cause diarrhea, fever, and abdominal cramps, and severe cases can even lead to death (CDC, 2012c).

Current methods for evaluating recreational and drinking water safety typically rely on the enumeration of FIB (U.S. Environmental Protection Agency [EPA], 2012). FIB are used to indicate the potential for illness associated with fecal contamination (EPA, 2012). Primary contact refers to water-contact activities where immersion and ingestion are likely, such as swimming, water skiing, and diving. Even though most tributaries are not used for “primary contact” recreational

water activities, many of these major tributaries enter the Great Lakes at or near a beach which is designated as a recreation area. Most tributaries are used for partial, or secondary, contact recreation, indicating water activities where there is the potential of submergence, such as boating and fishing. FIB concentrations and the occurrence of bacterial pathogens associated with these bodies of water are important, not only to partial contact recreational users in rivers but ultimately to primary contact recreational users at Great Lakes beaches. The partial body contact recreation standard (Dufour and Ballantine, 1986) was used for data interpretation for this report because the EPA 2012 guidance does not include a numerical standard for partial body contact in recreational waters.

### Purpose and Scope

This report presents data obtained during a Great Lakes-wide study conducted in 2011 by the USGS as part of the GLRI. The purpose of this study was to determine the occurrence and distribution of FIB and pathogenic gene markers within 22 major tributaries to the Great Lakes. Normal-flow and high-flow samples were analyzed for FIB and for pathogens *Shigella*, *Campylobacter* (*jejuni* and *coli*), *Salmonella*, and STEC, which included *E. coli* O157:H7. The relations of FIB concentrations and bacterial pathogen gene occurrence to recreational water quality criteria were also assessed.

### Description of Study Area

Approximately 132,208 miles of tributaries (EPA and USGS, 2012), with a drainage area of almost 295,000 square miles (Great Lakes Information Network, 2013), drain into the Great Lakes from six states (Indiana, Michigan, Minnesota, New York, Ohio, Wisconsin) and Canada. The 22 monitoring locations are grouped on the basis of predominant land cover surrounding the major tributaries in this study (fig. 1) (USGS, 2011) and were distributed geographically throughout the Great Lakes region of the United States (fig. 2).

## Sampling and Analytical Methods

### Site Selection

All sampling locations were at USGS streamgaging stations. Twenty-two sampling locations (fig. 2, table 1) were selected (out of 59 GLRI Loadings study sites) to create a geographically distributed study and to address the variability in land cover throughout the Great Lakes region. Sampling locations, as well as timing of sample collection, were designed to complement the GLRI Loadings study (USGS, 2014). In addition to those sites selected from the GLRI Loadings study, the Paw Paw River at Riverside, MI, was included as a sampling location for this study because the site was routinely visited as part of another USGS MI-WSC project.

### Sample Collection

Hydrologic conditions were determined on the basis of mean daily discharge<sup>1</sup> at each USGS streamgauge on the date of collection, calculated from date of first record to current. Conditions were evaluated such that if daily discharge<sup>2</sup> was greater than or equal to the 75th percentile, the corresponding sample was considered a high-flow sample; otherwise, the sample was considered a normal-flow sample. Owing to a short period of record, the Grand River at Grand Rapids station was used to estimate hydrologic conditions for the Grand River at Eastmanville station. Water samples were collected during normal-flow conditions from March through October 2011 in an effort to target the recreational water season. Samples were also collected during high-flow conditions (storm events), but sampling frequency varied among sites in response to the frequency of storm events and availability of staff (table 1).

The sampling frequency depended on the sampling schedule of the GLRI Loadings study. Some sites were sampled more often than others, including those sites targeted for analysis of certain chemical constituents (such as wastewater indicator compounds) in addition to FIB and pathogenic gene markers.

Normal-flow and high-flow sampling events typically coincided with GLRI Loadings study sampling. All samples were collected by USGS personnel using either equal-width-increment (EWI) or multiple vertical sampling techniques designed to account for the variable distribution of constituents across the stream channel and throughout the water column (USGS, 2006). EWI samples were typically collected off bridges by using a crane apparatus with an attached DH-95 depth integrating sampler, or from boats. There were instances when wading measurements were made, but typically only during normal-flow conditions.

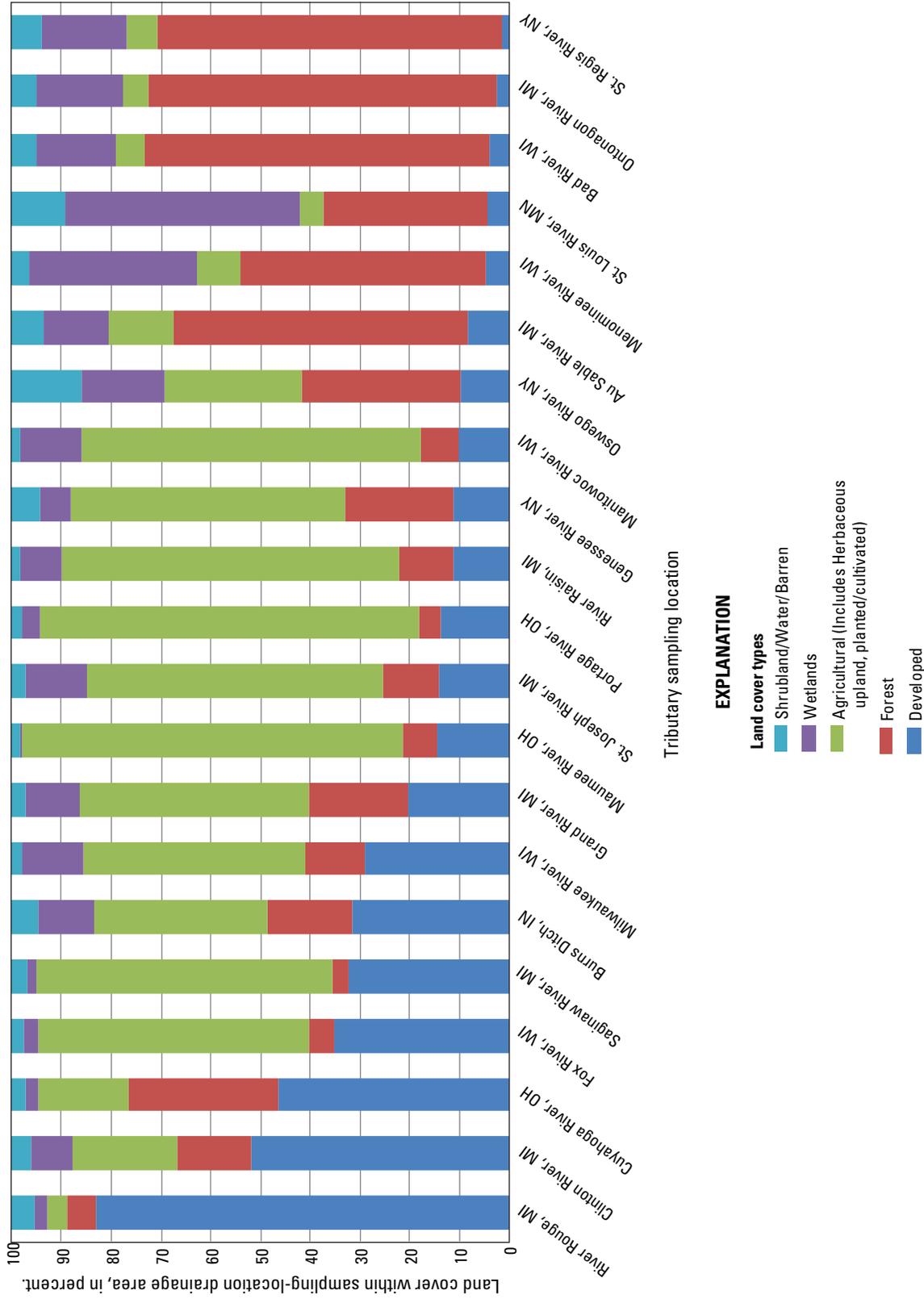
For each sample, a 1-liter sterile high density polyethylene (HDPE) sample bottle was filled to the shoulder to allow some head space for sample mixing. Once collected, water samples were placed on ice in a cooler (away from ultraviolet light) and shipped overnight to the MI-BaRL (Michigan Bacteriological Research Laboratory) at the MI-WSC in Lansing, Michigan. Upon receipt of the shipments, sample information and streamflow condition were logged in to the laboratory database system, and samples were placed into the refrigerator at 4–7 degrees Celsius (°C) until processing and analysis (typically the same day).

A total of 177 water samples were collected, including field quality-control (QC) samples. There were 59 normal-flow samples and 54 high-flow samples used for data analysis. Quality-control samples included 21 field blank samples, 21 source solution blank samples, and 22 field replicate samples.

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<sup>1</sup>The arithmetic mean of discharges for all instances of a particular calendar day on record or for a specific set of years.

<sup>2</sup>The arithmetic mean discharge for a single day.



**Figure 1.** Land cover surrounding tributary sites sampled as part of the Great Lakes Restoration Initiative (GLRI) pathogens study (data from 2006 National Land Cover Data [USGS, 2011]).

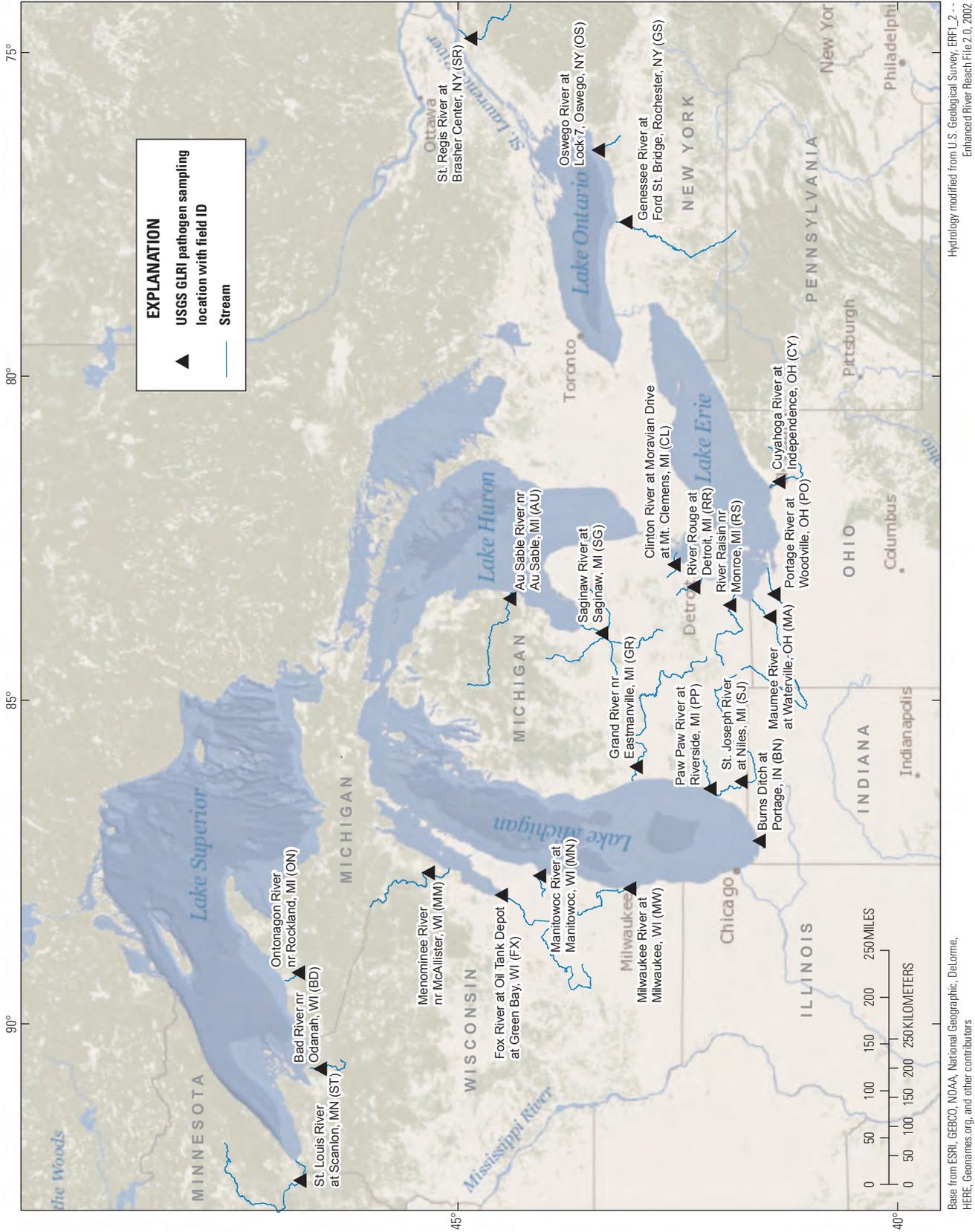


Figure 2. Great Lakes Restoration Initiative (GLRI) USGS tributary sampling locations and number of samples collected for pathogens study, 2011.

Field blanks, source solution blanks, and field replicate samples were collected according to the methods described in the USGS National Field Manual, with inorganic-grade reagent water being used as the sample medium (USGS, 2006). Twelve samples were excluded from the data analysis because the samples exceeded the holding time (48 hours) established

by the MI-BaRL for FIB culturing for nonregulatory purposes. There were also three instances where the bacteria plates were uncountable (coded as “unc”), owing to smearing or a heavy amount of particulates on the filter. A total of 165 water samples were included in the final analysis.

**Table 1.** Great Lakes Restoration Initiative (GLRI) pathogens study sampling locations, 2011.

[nr, near]

USGS site name	USGS site identification number	Field ID	Number of samples collected during normal flow	Number of samples collected during high flow
Indiana				
Burns Ditch at Portage	04095090	BN	3	4
Michigan				
Au Sable River nr Au Sable	04137500	AU	5	1
Clinton River at Moravian Drive at Mt. Clemens	04165500	CL	0	2
Grand River nr Eastmanville	04119400	GR	0	5
Ontonagon River nr Rockland	04040000	ON	8	1
Paw Paw River at Riverside	04102500	PP	2	5
River Rouge at Detroit	04166500	RR	1	2
River Raisin nr Monroe	04176500	RS	2	0
Saginaw River at Saginaw	04157000	SG	5	3
St. Joseph River at Niles	04101500	SJ	3	4
Minnesota				
St. Louis River at Scanlon	04024000	ST	7	2
New York				
Genesee River at Ford St. Bridge at Rochester	04231600	GS	3	3
Oswego River at Lock 7 at Oswego	04249000	OS	4	4
St. Regis River at Brasher Center	04269000	SR	5	5
Ohio				
Cuyahoga River at Independence	04208000	CY	1	4
Maumee River at Waterville	04193500	MA	2	1
Portage River at Woodville	04195500	PO	0	3
Wisconsin				
Bad River nr Odanah	04027000	BD	3	1
Fox River at Oil Tank Depot at Green Bay	040851385	FX	1	1
Menominee River nr McAllister	04067500	MM	2	0
Manitowoc River at Manitowoc	04085427	MN	1	1
Milwaukee River at Milwaukee	04087170	MW	1	1

## Microbiological Analyses

### Bacteria Enumeration, Enrichment, and Preservation

Water samples were analyzed for the presence of FIB and FIB concentrations by using standard membrane filtration and serial dilution methods (Myers and others, 2014). Fecal coliform bacteria were analyzed according to standard methods of the American Public Health Association and others (1998), *E. coli* bacteria were analyzed according to EPA method 1603 (EPA, 2009a), and enterococci bacteria were analyzed according to EPA method 1600 (EPA, 2009b). Media were prepared according to manufacturer's instructions.

A volume of 50 milliliters (mL) of sample water was filtered through a 0.45-micrometer-pore-size nylon-membrane filter (Advantec, USA) by means of aseptic techniques. The enriched filter was then placed on a selective growth medium for enumeration and enrichment of target bacterial pathogen groups and incubated at the optimal temperature (table 2).

The filters with *E. coli*, enterococci, and fecal coliform bacteria growth were transferred to a vial containing 1 mL of phosphate buffered saline solution (PBS), preserved with 20 percent glycerol, and frozen at  $-70^{\circ}\text{C}$  until further analysis; these samples are referred to as "glycerol stocks." The filters with *Campylobacter* and *Salmonella* growth were aseptically removed from the Gram Negative and Bolton Broth enrichments, and the remaining culture was centrifuged to form a pellet. The supernatant was decanted, and the pellet was resuspended in 1 mL of PBS solution and preserved with 20 percent glycerol and frozen at  $-70^{\circ}\text{C}$  until deoxyribonucleic acid (DNA) extraction.

### DNA Extraction

Glycerol stocks were thawed at room temperature and DNA was extracted by using the Qiagen QIAamp® DNA Mini Kit (Qiagen, USA), following the manufacturers protocol. DNA concentrations and purity were determined with a NanoDrop spectrophotometer (Thermo Scientific, USA). DNA was then stored at  $-20^{\circ}\text{C}$  until needed for Polymerase Chain Reaction (PCR) analysis for the selected gene targets (table 3).

### Polymerase Chain Reaction Assays for Pathogen Detection

Samples were analyzed by using PCR to determine the occurrence of pathogen gene markers for *Shigella sonnei*, *Campylobacter jejuni* and *coli*, *Salmonella*, and STEC. The primer sequences, positive controls, reaction details, and quality-control procedures used for the detection of the pathogens by PCR listed in table 3 were based on previously published studies (Haack and others, 2013).

All PCR assays were performed by using 1 microliter of DNA solution, representing 0.1–85 nanograms of template DNA, which was isolated from the appropriate growth enrichment. Results from these analyses indicated whether the target genes were present or absent in the sample above the method detection limit; however, PCR does not quantify the genes present. In order for the gene to be detected, the organism had to grow on the appropriate medium, and therefore must have been viable in the sample.

Laboratory filtration blanks were processed daily with every set of samples on all growth media and enrichments to ensure that the laboratory equipment was sterile and that aseptic laboratory techniques were being followed. One set of laboratory replicates was processed weekly on all growth media and enrichments to quantify method variability. If contamination was found in the laboratory blanks, either sample results were discarded or samples were rerun.

Standard quality-assurance and quality-control procedures were followed for all PCR reactions (EPA, 2004). Detection limits for all PCR reactions were determined by using serial dilution of DNA from control organisms. For approximately every 20 samples of any given PCR assay, PCR positive controls (DNA extracted from bacteria known to contain the target gene) and PCR negative controls (no template reactions) were included.

## Results of Microbiological Analyses

Of the 177 water samples collected, 12 samples were omitted from the data analysis because of holding-time violations and 64 of the samples were collected for the analysis of quality control. The 165 samples included in the analyses were assessed for occurrence and distribution of FIB during normal and high flows and for whether the samples exceeded the water quality criteria. These samples were also analyzed for specific pathogen genes by using PCR. Fecal indicator bacteria results are included in the USGS National Water Information System (NWIS) database.

### Occurrence and Distribution of Fecal Indicator Bacteria

Fecal coliform concentrations ranged from  $< 2$  to 950 colony-forming units per 100 mL (CFU/100 mL), *E. coli* concentrations ranged from  $< 2$  to 26,000 CFU/100 mL, and enterococci concentrations ranged from  $< 2$  to 31,000 CFU/100 mL. Fecal indicator bacteria concentrations for all samples are presented in appendix 1.

The lowest *E. coli* concentrations were recorded at the Au Sable River near Au Sable, MI (AU) (during high and normal flows), and the St. Joseph River at Niles, MI (during normal flow) (SJ). The highest *E. coli* concentrations were recorded at

the River Rouge at Detroit, MI (during high flow) (RR). The lowest enterococci concentrations were recorded at the Au Sable River near Au Sable, MI (AU) (during high and normal flows), and the Oswego River at Lock 7 at Oswego, NY (OS) (during high flow). The highest enterococci concentrations were recorded at the River Rouge at Detroit, MI (RR) (during high flow). The lowest *E. coli* and enterococci concentrations were found at sites dominated by forest and agricultural land cover, whereas the highest *E. coli* and enterococci concentrations were recorded at sites dominated by urban development.

Enterococci and *E. coli* concentrations were evaluated on the basis of whether the data met or exceeded the 1986 EPA criterion for moderate full body contact recreation for freshwater for a single sample (Dufour and Ballantine, 1986). The moderate full body contact recreation criterion was

considered most applicable for this study given the type of recreation typically found on rivers; for example, kayaking, canoeing, and fishing. The moderate full body contact criterion for enterococci for a single sample is greater than or equal to 78 CFU/100 mL; and for *E. coli*, the recreation criterion is greater than or equal to 298 CFU/100 mL for a single sample. Of 112 samples analyzed for FIB concentrations, 21 percent of samples exceeded the EPA recreation criterion for *E. coli*, and 40 percent of samples exceeded the criterion for enterococci.

During high-flow events, 43 percent of all samples exceeded the EPA recreation criterion for *E. coli*, and 61 percent of samples exceeded the criterion for enterococci. During normal-flow conditions, about 2 percent of samples exceeded the criterion for *E. coli*, and about 21 percent of samples exceeded the criterion for enterococci.

**Table 2.** Growth media, target organisms, and incubation temperature requirements for the Great Lakes Restoration Initiative (GLRI) pathogens study.

[mL, milliliters; °C, degrees Celsius; *E. coli*, *Escherichia coli*]

Growth media	Organism or method	Volume filtered (mL)	Incubation temperature and time requirement
mFC	Fecal coliform bacteria ( <i>E. coli</i> , <i>Shigella</i> )	50	44.5 °C ± 0.5 °C for 22–24 hours
Modified mTEC	<i>E. coli</i>	50, 10, 1	35–37 °C for 2 hours, then 44.5 °C ± 0.5 °C for 20–22 hours
mEI	Enterococci	50, 10, 1	41 °C ± 0.5 °C for 22–24 hours
Bolton Broth (BB) with Preston <sup>1</sup> supplement	<i>Campylobacter</i>	50	37 °C for 4 hours, then 42 °C for 40–44 hours
Gram Negative Broth (GN)	<i>Salmonella</i>	50	37°C ± 1.0 °C for 22–24 hours

<sup>1</sup>Oxoid, Cambridge, United Kingdom.

**Table 3.** Selected pathogen gene targets and virulence traits of the organism.

[*E. coli*, *Escherichia coli*]

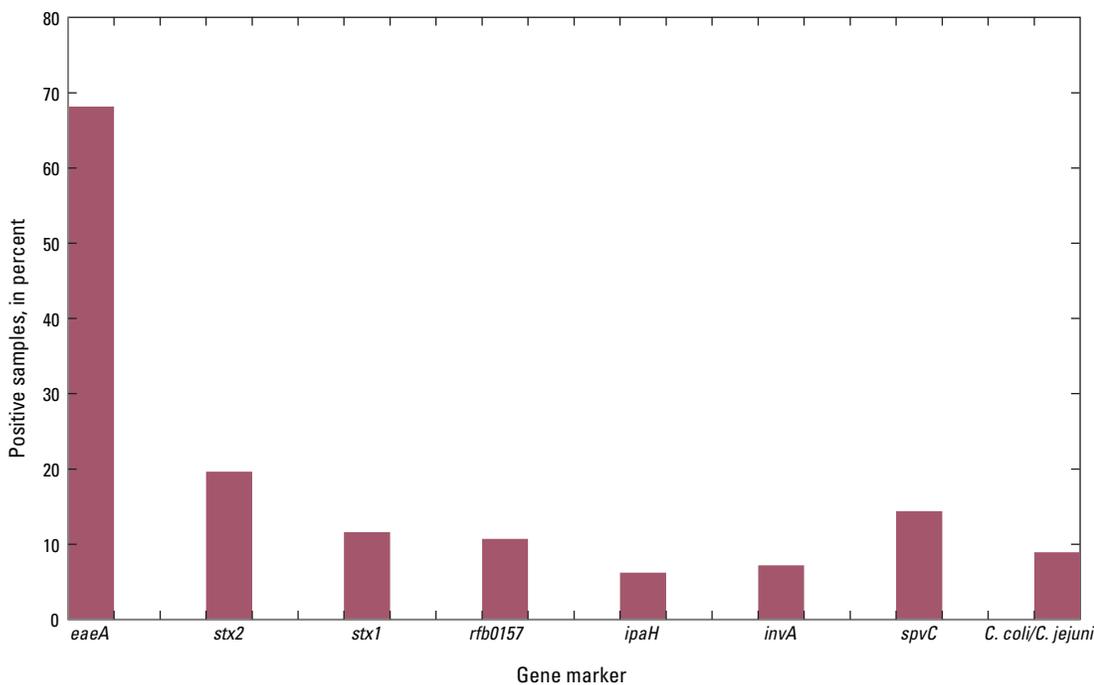
Pathogen	Gene target	Virulence trait
<i>E. coli</i>	<i>eaeA</i>	Attachment
<i>E. coli</i>	<i>stx2</i>	Severe toxin
<i>E. coli</i>	<i>stx1</i>	Moderate toxin
<i>E. coli</i>	<i>rfbO157</i>	Common outbreak strain
<i>Shigella</i>	<i>ipaH</i>	Invasion
<i>Salmonella</i>	<i>invA</i>	Invasion
<i>Salmonella</i>	<i>spvC</i>	Multiple virulence traits
<i>Campylobacter jejuni</i> and <i>coli</i>	<i>16s rDNA</i> (Campy)	Pathogenic species marker

## Occurrence and Distribution of Bacterial Pathogen Gene Markers

Data from the *E. coli* pathogen analysis showed that 68 percent of samples contained the *eaeA* gene, 20 percent of samples contained the *stx2* gene, 12 percent of samples contained the *stx1* gene, and 11 percent of samples contained the *rfbO157* gene (fig. 3). Data from the *Shigella* analysis showed that 6 percent of samples contained the *ipaH* gene. Data from the *Salmonella* analysis showed that 7 percent of samples contained the *invA* gene and 14 percent of samples contained the *spvC* gene. Data from the *Campylobacter* analysis showed that 9 percent of samples contained the 16s rDNA gene (fig. 3).

Detections of bacterial pathogen gene markers by site and by sample are listed in appendix 3.

The frequency of pathogen gene markers detected in each sample is summarized in table 4. The greatest number of pathogen gene markers detected in any one sample was seven (River Rouge at Detroit, MI, during a May 26, 2011, high-flow event). This same May 26, 2011 sample from the River Rouge also contained the highest *E. coli* and enterococci concentrations of samples collected during this project (*E. coli* value of 26,000 CFU/100 mL; enterococci value, 31,000 CFU/100 mL).



**Figure 3.** Frequency of samples testing positive for pathogen gene markers (*C. coli*, *Campylobacter coli*; *C. jejuni*, *Campylobacter jejuni*).

**Table 4.** Summary of the frequency of pathogen gene detections in a single sample.

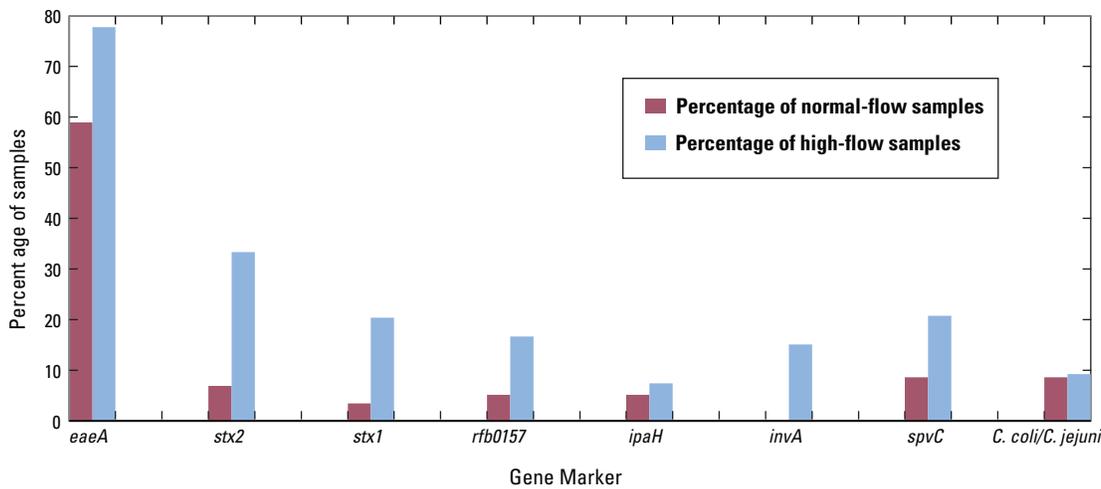
[<, less than]

Flow conditions	Number of bacterial pathogen genes present									
	0	1	2	3	4	5	6	7	8	
	Percentage of samples in which the indicated number of genes were detected									
High flow	10	12	13	2	3.5	6	0	1	0	
Normal flow	16	25	10	1	<1	0	0	0	0	
Total percent detection	26	37	23	3	4	6	0	1	0	

For those samples in which 3 or more genes were present, 14 of the 16 samples were collected during high-flow conditions.

Pathogen frequency was analyzed in relation to hydro-logic condition among all sites, as illustrated in figure 4. Overall, a higher percentage of samples tested positive for pathogens under high-flow conditions than during normal-flow conditions. For *Campylobacter*, the differences in the percentage of samples that were positive for pathogens between high- and normal-flow conditions were less obvious. The *invA* gene for *Salmonella* was found only during high flows.

The frequencies of pathogen gene occurrence in samples that met and exceeded the EPA criteria for moderate full body contact recreation for *E. coli* and enterococci were compared for the 22 sites (table 5). Fisher’s Exact Test was run to determine if there was a significant difference (p-value less than 0.05) between gene detection in samples that met or exceeded the water quality criteria. Overall, there was a greater occurrence of pathogen gene markers in samples that exceeded the moderate full body contact recreation criteria for both *E. coli* and enterococci, the exception being a greater occurrence of the *ipaH* gene marker for *Shigella* in samples that met the criterion for *E. coli*.



**Figure 4.** Percentage of pathogen gene markers in relation to water quality criteria (*C. coli*, *Campylobacter coli*; *C. jejuni*, *Campylobacter jejuni*).

**Table 5.** Percentage of samples that tested positive for pathogenic gene markers during high-flow and normal-flow conditions.

[*C. coli*, *Campylobacter coli*; *C. jejuni*, *Campylobacter jejuni*, *E. coli*, *Escherichia coli*; n, number of samples; <, less than; nsd, no significant difference; p-value, p-value less than 0.05 = significant difference between groups, Fisher’s Exact Test]

Water quality criteria <sup>1</sup> status	n	<i>eaeA</i>	<i>stx2</i>	<i>stx1</i>	<i>rfb0157</i>	<i>ipaH</i>	<i>invA</i>	<i>spvC</i>	<i>C. coli/C. jejuni</i>
Meet <i>E. coli</i> criterion	88	60	11	3	5	7	0	9	6
Exceed <i>E. coli</i> criterion	24	96	50	42	33	4	33	33	21
<b>p-value</b>	—	<0.05	<0.05	<0.05	<0.05	nsd	<0.05	<0.05	<0.05
Meet enterococci criterion	67	54	10	5	3	6	0	3	2
Exceed enterococci criterion	45	91	33	22	22	7	18	31	20
<b>p-value</b>	—	<0.05	<0.05	<0.05	<0.05	nsd	<0.05	<0.05	<0.05

<sup>1</sup>Water quality criteria = EPA Water Quality Criteria for Moderate Full Body Contact (Dufour and Ballantine, 1986).

## Quality-Control Results

To assess for bias and precision, 21 field blanks, 21 source solution blanks, and 22 field replicates were collected and processed in the same manner as the regular field samples. All field blanks and source solution water blanks were negative (no growth) for all FIB except for a sample collected at the River Raisin on August 30, 2011, which had an enterococci count of 2E (appendix 2). For PCR analysis, all no-template controls (negative controls) were found to be negative, and all positive assay controls yielded appropriate responses. However, the *eaeA*, *rfb0157*, and *ipaH* genes were infrequently detected in the field blanks and source solution blanks, most likely as a result of contamination in the source solution blank water (appendix 4). There were two dates—September 7, 2011, and August 24, 2011—for which environmental samples were collected in association with the source solution blank and field blank samples where there may have been possible contamination. The environmental sample, source solution blank, and field blank samples from these dates have been qualified with a “V” (analyte was detected in both the environmental sample and the associated blanks) as a result of possible contamination and data were not included in analyses (appendixes 3 and 4).

For all FIB field replicates, a relative standard deviation (RSD) was calculated and a mean RSD was computed (table 6) by using the formula below:

$$S = \frac{\sqrt{(X1 - \bar{X})^2 + (X2 - \bar{X})^2 + (X3 - \bar{X})^2 + \dots}}{n - 1}$$

**Table 6.** Relative standard deviation of field replicates for quantitative microbial methods in the Great Lakes Restoration Initiative (GLRI) pathogens study.

[*E. coli*, *Escherichia coli*; RSD, relative standard deviation]

Statistic	<i>E. coli</i>	Enterococci	Fecal coliforms
Average RSD, in percent	25.3	28.4	31.1
Number of field replicates	22	22	22

**Table 7.** Frequency of agreement between field replicates for qualitative microbial methods in the Great Lakes Restoration Initiative (GLRI) pathogens study.

[*C. coli*, *Campylobacter coli*; *C. jejuni*, *Campylobacter jejuni*]

Statistic	Pathogen gene markers							<i>C. coli/C. jejuni</i>
	<i>eaeA</i>	<i>stx2</i>	<i>stx1</i>	<i>rfb0157</i>	<i>ipaH</i>	<i>invA</i>	<i>spvC</i>	
Percent agreement	82	95	95	82	100	95	91	95
Number of field replicates	22	22	22	22	22	22	22	22

In all cases, the average RSD fell below the recommended 36 percent according to the initial and ongoing precision and recovery acceptance criteria that were used as a benchmark for replicate environmental samples (table 6) (EPA, 2009a).

For pathogen analysis, which yields only a presence or absence result, the frequencies of agreement between replicates were determined by dividing the number of paired replicates in agreement by the total number of replicates (table 7).

The field blank and source solution blank data results are presented in appendixes 2 and 4. All replicate data are presented in appendixes 1 and 3, along with the associated environmental sample.

## Summary

A total of 177 environmental samples were collected at USGS streamgaging stations during both normal-flow and high-flow conditions in 22 tributaries to the Great Lakes and were analyzed for the presence of FIB concentrations and to determine the occurrence of pathogen gene markers for *Shigella* species, *Campylobacter jejuni* and *coli*, *Salmonella* species, and pathogenic *E. coli*, including STEC. Results from a total of 165 samples were evaluated because 12 samples did not meet quality-control measures.

*E. coli* concentrations ranged from less than 2 to 26,000 CFU/100 mL. Enterococci concentrations ranged from less than 2 to 31,000 CFU/100 mL. Fecal coliform bacteria concentrations ranged from less than 2 to 950 CFU/100 mL. Data from *E. coli* pathogen analyses showed that 68 percent of samples contained the *eaeA* gene, 20 percent of samples contained the *stx2* gene, 12 percent of samples contained the *stx1* gene, and 11 percent of samples contained the *rfb0157* gene. Data from the *Shigella* analysis showed that 6 percent of samples contained the *ipaH* gene. Data from the *Salmonella* analysis showed that 7 percent of samples contained the *invA* gene and 14 percent of samples contained the *spvC* gene. Data from the *Campylobacter* analysis showed that 9 percent of samples contained the 16S rDNA gene of *Campylobacter coli* and *Campylobacter jejuni*.

There were no samples in which all eight pathogen gene markers were detected; however, 88 percent of the samples that had three or more pathogen gene markers present were collected during high-flow conditions. Overall, there were a higher percentage of samples that tested positive for pathogens during high-flow conditions than normal-flow conditions, and there was a greater occurrence of pathogen gene markers in samples that exceeded the EPA moderate full body contact recreation criteria for *E. coli* and enterococci (Dufour, 1986) than in samples that met the EPA criteria; the exception was a greater occurrence of the *ipaH* gene marker for *Shigella* in samples that met the EPA criterion for *E. coli* than in samples that exceeded the EPA criterion for *E. coli*.

## Acknowledgements

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# Appendixes

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1. Fecal indicator bacteria concentrations in Great Lakes tributaries, 2011 .....14
2. Quality-control data—Fecal indicator bacteria concentrations in Great Lakes tributaries, 2011 .....20
3. Pathogen gene detection in Great Lakes tributaries, 2011 .....22
4. Quality-control data—Pathogen gene detection in Great Lakes tributaries, 2011 .....28

**Appendix 1.** Fecal indicator bacteria concentrations in Great Lakes tributaries, 2011.

[Streamgaging stations corresponding to station IDs and field IDs are listed in table 1 in the main report. USGS, U.S. Geological Survey; Assoc. environ. sample, associated environmental sample; *E. coli*, *Escherichia coli*; CFU/100 mL, colony-forming units per 100 milliliters; ns, no sample; unc, uncountable; E, estimated value; <, less than; >, greater than]

USGS station ID	Field ID	Sample collection date	Flow conditions	Sample type	Assoc. environ. sample	Fecal coliforms (CFU/100mL)	Remark code	<i>E. coli</i> (CFU/100 mL)	Remark code	Enterococci (CFU/100 mL)	Remark code
04137500	AU-01	3/15/2011	Normal	Regular		2	<	2	<	2	<
04137500	AU-04	6/15/2011	Normal	Regular		2	E	2	E	2	<
04137500	AU-05	7/13/2011	Normal	Regular		74	<	2	<	2	<
04137500	AU-06	7/13/2011	Normal	Replicate	AU-05	34	E	2	E	2	E
04137500	AU-07	8/23/2011	Normal	Regular		2	E	2	<	4	E
04137500	AU-10	9/13/2011	Normal	Regular		4	E	2	E	2	<
04027000	BD-04	7/12/2011	Normal	Regular		84		44		23	E
04027000	BD-05	7/12/2011	Normal	Replicate	BD-04	58		52		18	E
04027000	BD-08	8/16/2011	Normal	Regular		160	E	220		25	E
04027000	BD-09	9/20/2011	Normal	Regular		unc		66		28	E
04095090	BN-01	3/30/2011	Normal	Regular		22	E	8	E	8	E
04095090	BN-05	7/12/2011	Normal	Regular		120	>	140		140	E
04095090	BN-08	8/11/2011	Normal	Regular		430	E	140		120	
04095090	BN-09	8/11/2011	Normal	Replicate	BN-08	340	E	150		140	E
04208000	CY-06	8/31/2011	Normal	Regular		unc		220		150	E
040851385	FX-02	8/16/2011	Normal	Regular		130	E	23	E	8	E
040851385	FX-03	8/16/2011	Normal	Replicate	FX-02	110		30	E	13	E
04231600	GS-01	4/5/2011	Normal	Regular		120	>	520		230	E
04231600	GS-09	7/18/2011	Normal	Regular		180	E	6	E	2	E
04231600	GS-10	7/18/2011	Normal	Replicate	GS-09	190	E	2	E	2	E
04231600	GS-13	8/8/2011	Normal	Regular		84		32	E	13	E
04193500	MA-02	4/19/2011	Normal	Regular		42		88		76	
04193500	MA-03	7/27/2011	Normal	Regular		120	>	18	E	20	E
04193500	MA-04	7/27/2011	Normal	Replicate	MA-03	120	>	26	E	390	
04067500	MM-02	7/7/2011	Normal	Regular		100		10	E	690	E
04067500	MM-03	9/7/2011	Normal	Regular		6	E	8	E	7	E

## Appendix 1. Fecal indicator bacteria concentrations in Great Lakes tributaries, 2011.—Continued

[Streamgaging stations corresponding to station IDs and field IDs are listed in table 1 in the main report. USGS, U.S. Geological Survey; Assoc. environ. sample, associated environmental sample; *E. coli*, *Escherichia coli*; CFU/100 mL, colony-forming units per 100 milliliters; ns, no sample; unc, uncountable; E, estimated value; <, less than; >, greater than]

USGS station ID	Field ID	Sample collection date	Flow conditions	Sample type	Assoc. environ. sample	Fecal coliforms (CFU/100mL)	Remark code	<i>E. coli</i> (CFU/100 mL)	Remark code	Enterococci (CFU/100 mL)	Remark code
04067500	MM-04	9/7/2011	Normal	Replicate	MM-03	20	E	2	E	6	E
04085427	MN-03	9/8/2011	Normal	Regular		72		68		140	E
04085427	MN-04	9/8/2011	Normal	Replicate	MN-03	94		120		140	E
04087170	MW-02	9/20/2011	Normal	Regular		48		30	E	8	E
04087170	MW-03	9/20/2011	Normal	Replicate	MW-02	50		39	E	5	E
04040000	ON-01	3/23/2011	Normal	Regular		40		50		72	
04040000	ON-02	4/5/2011	Normal	Regular		28	E	35	E	94	
04040000	ON-03	4/26/2011	Normal	Regular		6	E	6	E	7	E
04040000	ON-04	5/25/2011	Normal	Regular		52		74		44	
04040000	ON-05	6/6/2011	Normal	Regular		10	E	5	E	4	E
04040000	ON-07	7/14/2011	Normal	Regular		340	E	30	E	58	
04040000	ON-08	8/17/2011	Normal	Regular		16	E	10	E	38	E
04040000	ON-09	9/13/2011	Normal	Regular		38	E	35	E	27	E
04040000	ON-10	9/13/2011	Normal	Replicate	ON-09	32	E	15	E	20	E
04249000	OS-02	4/7/2011	Normal	Regular		30	E	22	E	4	E
04249000	OS-05	6/8/2011	Normal	Regular		86		68		43	E
04249000	OS-10	7/18/2011	Normal	Regular		150	E	33	E	76	
04249000	OS-11	7/18/2011	Normal	Replicate	OS-10	120	>	140		220	
04249000	OS-14	8/8/2011	Normal	Regular		12	E	20	E	7	E
04102500	PP-01	4/11/2011	Normal	Regular		42		82		38	E
04102500	PP-06	9/8/2011	Normal	Regular		170	E	72		120	
04166500	RR-03	8/17/2011	Normal	Regular		120	>	140		320	
04166500	RR-04	8/17/2011	Normal	Replicate	RR-03	420	E	430		440	
04176500	RS-01	6/16/2011	Normal	Regular		100		110		66	
04176500	RS-02	8/23/2011	Normal	Regular		52		20	E	460	
04176500	RS-03	8/23/2011	Normal	Replicate	RS-02	44		44		370	

**Appendix 1.** Fecal indicator bacteria concentrations in Great Lakes tributaries, 2011.—Continued

[Streamgaging stations corresponding to station IDs and field IDs are listed in table 1 in the main report. USGS, U.S. Geological Survey; Assoc. environ. sample, associated environmental sample; *E. coli*, *Escherichia coli*; CFU/100 mL, colony-forming units per 100 milliliters; ns, no sample; unc, uncountable; E, estimated value; <, less than; >, greater than]

USGS station ID	Field ID	Sample collection date	Flow conditions	Sample type	Assoc. environ. sample	Fecal coliforms (CFU/100mL)	Remark code	<i>E. coli</i> (CFU/100 mL)	Remark code	Enterococci (CFU/100 mL)	Remark code
04157000	SG-01	3/15/2011	Normal	Regular		210	E	66	E	160	E
04157000	SG-05	6/16/2011	Normal	Regular		150	E	36	E	42	E
04157000	SG-06	7/14/2011	Normal	Regular		950	E	25	E	12	E
04157000	SG-07	7/14/2011	Normal	Replicate	SG-06	850	E	20	E	13	E
04157000	SG-08	8/23/2011	Normal	Regular		52		30	E	12	E
04157000	SG-11	9/14/2011	Normal	Regular		110		60		40	
04101500	SJ-02	4/7/2011	Normal	Regular		26	E	2	<	7	E
04101500	SJ-07	7/27/2011	Normal	Regular		110		74		52	
04101500	SJ-08	8/24/2011	Normal	Regular		370	E	130		52	
04269000	SR-01	3/24/2011	Normal	Regular		16	E	6	E	2	E
04269000	SR-04	4/26/2011	Normal	Regular		14	E	8	E	2	E
04269000	SR-07	6/21/2011	Normal	Regular		ns		ns		ns	
04269000	SR-08	7/26/2011	Normal	Regular		170	E	130		46	
04269000	SR-09	7/26/2011	Normal	Replicate	SR-08	210	E	140		52	
04269000	SR-13	9/20/2011	Normal	Regular		150	E	180	E	110	
04024000	ST-01	3/15/2011	Normal	Regular		8	E	2	E	2	E
04024000	ST-02	4/20/2011	Normal	Regular		2	E	4	E	5	E
04024000	ST-04	6/14/2011	Normal	Regular		12	E	3	E	2	E
04024000	ST-05	6/23/2011	Normal	Regular		26	E	18	E	44	E
04024000	ST-06	7/21/2011	Normal	Regular		120		26	E	28	E
04024000	ST-07	7/21/2011	Normal	Replicate	ST-06	110		42		42	
04024000	ST-11	8/31/2011	Normal	Regular		74		40		20	E
04024000	ST-12	9/13/2011	Normal	Regular		42		25	E	10	E
04137500	AU-02	4/12/2011	High	Regular		2	<	2	<	2	<
04027000	BD-01	4/11/2011	High	Regular		110		82	E	280	
04027000	BD-02	4/11/2011	High	Replicate	BD-01	140	E	56		270	

## Appendix 1. Fecal indicator bacteria concentrations in Great Lakes tributaries, 2011.—Continued

[Streamgaging stations corresponding to station IDs and field IDs are listed in table 1 in the main report. USGS, U.S. Geological Survey; Assoc. environ. sample, associated environmental sample; *E. coli*, *Escherichia coli*; CFU/100 mL, colony-forming units per 100 milliliters; ns, no sample; unc, uncountable; E, estimated value; <, less than; >, greater than]

USGS station ID	Field ID	Sample collection date	Flow conditions	Sample type	Assoc. environ. sample	Fecal coliforms (CFU/100mL)	Remark code	<i>E. coli</i> (CFU/100 mL)	Remark code	Enterococci (CFU/100 mL)	Remark code
04027000	BD-03	5/24/2011	High	Regular		340	E	510	E	150	E
04095090	BN-02	4/28/2011	High	Regular		120	>	1400	E	1100	E
04095090	BN-03	5/25/2011	High	Regular		610	E	710	E	350	E
04095090	BN-04	6/8/2011	High	Regular		46	E	17	E	7	E
04095090	BN-10	9/7/2011	High	Regular		94		48		56	
04165500	CL-01	5/16/2011	High	Regular		120	>	2500		3800	
04165500	CL-02	8/17/2011	High	Regular		230	E	200	E	100	E
04165500	CL-03	8/17/2011	High	Replicate	CL-02	160	E	140	E	84	E
04208000	CY-01	4/27/2011	High	Regular		490	E	590	E	250	E
04208000	CY-02	7/26/2011	High	Regular		120	>	180	>	100	>
04208000	CY-03	7/26/2011	High	Replicate	CY-02	120	>	250	>	78	>
04208000	CY-07	9/20/2011	High	Regular		120	>	6100	>	8000	E
04208000	CY-08	10/26/2011	High	Regular		430	E	430	E	420	E
040851385	FX-01	5/4/2011	High	Regular		14	E	64	E	4	E
04119400	GR-01	4/29/2011	High	Regular		310	E	290	E	280	E
04119400	GR-02	5/18/2011	High	Regular		140	E	230	E	34	E
04119400	GR-03	6/2/2011	High	Regular		70		48		17	E
04119400	GR-04	6/29/2011	High	Regular		240	E	82	E	12	E
04119400	GR-05	6/29/2011	High	Replicate	GR-04	270	E	80	E	8	E
04119400	GR-06	8/2/2011	High	Regular		130	E	90	E	17	E
04231600	GS-02	4/26/2011	High	Regular		400	E	320	E	270	E
04231600	GS-03	5/17/2011	High	Regular		120	>	1200	E	1400	E
04231600	GS-04	6/2/2011	High	Regular		unc		390		120	
04193500	MA-01	3/8/2011	High	Regular		570	E	650	E	980	E
04085427	MN-02	7/6/2011	High	Regular		410	E	70	E	50	E
04087170	MW-01	5/3/2011	High	Regular		28	E	58	E	27	E

**Appendix 1.** Fecal indicator bacteria concentrations in Great Lakes tributaries, 2011.—Continued

[Streamgaging stations corresponding to station IDs and field IDs are listed in table 1 in the main report. USGS, U.S. Geological Survey; Assoc. environ. sample, associated environmental sample; *E. coli*, *Escherichia coli*; CFU/100 mL, colony-forming units per 100 milliliters; ns, no sample; unc, uncountable; E, estimated value; <, less than; >, greater than]

USGS station ID	Field ID	Sample collection date	Flow conditions	Sample type	Assoc. environ. sample	Fecal coliforms (CFU/100mL)	Remark code	<i>E. coli</i> (CFU/100 mL)	Remark code	Enterococci (CFU/100 mL)	Remark code
04040000	ON-06	6/23/2011	High	Regular		120	>	920	E	570	
04249000	OS-01	3/9/2011	High	Regular		150	E	100		64	
04249000	OS-03	4/27/2011	High	Regular		70		88		66	
04249000	OS-04	5/9/2011	High	Regular		4	E	18	E	2	<
04249000	OS-15	9/7/2011	High	Regular		70		15	E	11	E
04195500	PO-01	3/8/2011	High	Regular		580	E	650		570	
04195500	PO-02	4/20/2011	High	Regular		120	>	1400	E	3600	
04195500	PO-03	7/27/2011	High	Regular		120	>	240		310	
04195500	PO-04	7/27/2011	High	Replicate	PO-03	120	>	220		280	
04102500	PP-02	4/21/2011	High	Regular		120	>	790		1200	E
04102500	PP-03	6/1/2011	High	Regular		96		100		72	
04102500	PP-04	7/19/2011	High	Regular		120	>	4800		6700	E
04102500	PP-05	8/3/2011	High	Regular		410	E	630		420	E
04102500	PP-08	10/4/2011	High	Regular		120		70		82	
04166500	RR-01	5/16/2011	High	Regular		120	>	2600		5700	
04166500	RR-02	5/26/2011	High	Regular		120	>	26000	E	31000	E
04157000	SG-02	4/6/2011	High	Regular		240	E	220		440	
04157000	SG-03	5/2/2011	High	Regular		100		190	E	98	
04157000	SG-04	5/16/2011	High	Regular		120	>	1200	E	960	E
04101500	SJ-01	3/9/2011	High	Regular		unc		96		440	
04101500	SJ-03	4/21/2011	High	Regular		120	>	860	E	1300	E
04101500	SJ-04	4/28/2011	High	Regular		120	>	1400	E	870	E
04101500	SJ-05	6/28/2011	High	Replicate	SJ-06	260	E	100		23	
04101500	SJ-06	6/28/2011	High	Regular		380	E	58		40	
04269000	SR-02	4/6/2011	High	Regular		16	E	5	E	14	E
04269000	SR-03	4/12/2011	High	Regular		50		40	E	33	E

**Appendix 1. Fecal indicator bacteria concentrations in Great Lakes tributaries, 2011.—Continued**

[Streamgaging stations corresponding to station IDs and field IDs are listed in table 1 in the main report. USGS, U.S. Geological Survey; Assoc. environ. sample, associated environmental sample; *E. coli*, *Escherichia coli*; CFU/100 mL, colony-forming units per 100 milliliters; ns, no sample; unc, uncountable; E, estimated value; <, less than; >, greater than]

USGS station ID	Field ID	Sample collection date	Flow conditions	Sample type	Assoc. environ. sample	Fecal coliforms (CFU/100mL)	Remark code	<i>E. coli</i> (CFU/100 mL)	Remark code	Enterococci (CFU/100 mL)	Remark code
04269000	SR-05	5/5/2011	High	Regular		24	E	46		17	E
04269000	SR-06	5/24/2011	High	Regular		30	E	60		30	E
04269000	SR-12	8/23/2011	High	Regular		220	E	220		210	
04024000	ST-03	5/1/2011	High	Regular		16	E	17	E	6	E
04024000	ST-10	8/3/2011	High	Regular		580	E	790		360	

**Appendix 2.** Quality-control data—Fecal indicator bacteria concentrations in Great Lakes tributaries, 2011.

[Streamgaging stations corresponding to station IDs and field IDs are listed in table 1 in the main report. USGS, U.S. Geological Survey; Assoc. environ. sample, associated environmental sample; *E. coli*, *Escherichia coli*; CFU/100 mL, colony-forming units per 100 milliliters; E, estimated value; <, less than; >, greater than; N/A, not applicable; ns, no sample]

USGS station ID	Field ID	Sample collection date	Sample type	Assoc. environ. sample	Fecal coliforms (CFU/100 mL)	Remark code	<i>E. coli</i> (CFU/100 mL)	Remark code	Enterococci (CFU/100 mL)	Remark code
04137500	AU-08	8/23/2011	Field blank	AU-07	2	<	2	<	2	<
04027000	BD-06	7/12/2011	Field blank	BD-04	2	<	2	<	2	<
04095090	BN-06	7/12/2011	Field blank	BN-05	2	<	2	<	2	<
04165500	CL-04	11/1/2011	Field blank	ns	2	<	2	<	2	<
04208000	CY-04	7/26/2011	Field blank	CY-02	2	<	2	<	2	<
040851385	FX-04	9/19/2011	Field blank	ns	2	<	2	<	2	<
04119400	GR-07	10/31/2011	Field blank	ns	2	<	2	<	2	<
04231600	GS-11	7/18/2011	Field blank	GS-09	2	<	2	<	2	<
04193500	MA-05	7/27/2011	Field blank	MA-03	2	<	2	<	2	<
04067500	MM-05	9/7/2011	Field blank	MM-03	2	<	2	<	2	<
04085427	MN-05	9/8/2011	Field blank	MN-03	2	<	2	<	2	<
04087170	MW-04	9/20/2011	Field blank	MW-02	2	<	2	<	2	<
04040000	ON-12	9/20/2011	Field blank	ns	2	<	2	<	2	<
04249000	OS-12	7/18/2011	Field blank	OS-10	2	<	2	<	2	<
04195500	PO-05	7/27/2011	Field blank	PO-03	2	<	2	<	2	<
04166500	RR-05	10/31/2011	Field blank	ns	2	<	2	<	2	<
04176500	RS-04	8/30/2011	Field blank	ns	2	<	2	<	2	E
04157000	SG-09	8/23/2011	Field blank	SG-08	2	<	2	<	2	<
04101500	SJ-09	8/24/2011	Field blank	SJ-08	2	<	2	<	2	<
04269000	SR-10	7/26/2011	Field blank	SR-08	2	<	2	<	2	<
04024000	ST-08	7/21/2011	Field blank	ST-06	2	<	2	<	2	<
04137500	AU-09	8/22/2011	Source solution blank	ns	2	<	2	<	2	<
04027000	BD-07	7/12/2011	Source solution blank	BD-04	2	<	2	<	2	<
04095090	BN-07	7/12/2011	Source solution blank	BN-05	2	<	2	<	2	<
04165500	CL-05	11/1/2011	Source solution blank	ns	2	<	2	<	2	<
04208000	CY-05	7/26/2011	Source solution blank	CY-02	2	<	2	<	2	<

**Appendix 2. Quality-control data—Fecal indicator bacteria concentrations in Great Lakes tributaries, 2011.—Continued**

[Streamgaging stations corresponding to station IDs and field IDs are listed in table 1 in the main report. USGS, U.S. Geological Survey; Assoc. environ. sample, associated environmental sample; *E. coli*, *Escherichia coli*; CFU/100 mL, colony-forming units per 100 milliliters; E, estimated value; <, less than; >, greater than; N/A, not applicable; ns, no sample]

USGS station ID	Field ID	Sample collection date	Sample type	Assoc. environ. sample	Fecal coliforms (CFU/100 mL)	Remark code	<i>E. coli</i> (CFU/100 mL)	Remark code	Enterococci (CFU/100 mL)	Remark code
040851385	FX-05	9/19/2011	Source solution blank	ns	2	<	2	<	2	<
04119400	GR-08	10/31/2011	Source solution blank	ns	2	<	2	<	2	<
04231600	GS-12	7/18/2011	Source solution blank	GS-09	2	<	2	<	2	<
04193500	MA-06	7/27/2011	Source solution blank	MA-03	2	<	2	<	2	<
04067500	MM-06	9/19/2011	Source solution blank	ns	2	<	2	<	2	<
04085427	MN-06	9/19/2011	Source solution blank	ns	2	<	2	<	2	<
04087170	MW-05	9/20/2011	Source solution blank	MW-02	2	<	2	<	2	<
04040000	ON-11	9/20/2011	Source solution blank	ns	2	<	2	<	2	<
04249000	OS-13	7/18/2011	Source solution blank	OS-10	2	<	2	<	2	<
04195500	PO-06	7/27/2011	Source solution blank	PO-03	2	<	2	<	2	<
04166500	RR-06	10/31/2011	Source solution blank	ns	2	<	2	<	2	<
04176500	RS-05	8/30/2011	Source solution blank	ns	2	<	2	<	2	<
04157000	SG-10	8/23/2011	Source solution blank	SG-08	2	<	2	<	2	<
04101500	SJ-10	8/24/2011	Source solution blank	SJ-08	2	<	2	<	2	<
04269000	SR-11	7/26/2011	Source solution blank	SR-08	2	<	2	<	2	<
04024000	ST-09	7/21/2011	Source solution blank	ST-06	2	<	2	<	2	<





**Appendix 3. Pathogen gene detection in Great Lakes tributaries, 2011.—Continued**

[Streamgaging stations corresponding to station IDs and field IDs are listed in table 1 in the main report. USGS, U.S. Geological Survey; Assoc. environ. sample, associated environmental sample; *E. coli*, *Escherichia coli*; +, gene present in sample; —, gene absent in sample; ns, no sample; V—remark code indicating possible contamination (analyte was detected in both the environmental sample and the associated blanks)]

USGS station ID	Field ID	Sample collection date	Flow conditions	Sample type	Assoc. environ. sample	E. coli gene data				Shigella gene data		Salmonella gene data		Campylobacter gene data	
						eaeA	stx2	stx1	rfb O157	ipaH	invA	spvC	C. coli/ C. jejuni		
04176500	RS-02	8/23/2011	Normal	Regular		+	—	—	—	+	—	—	—		
04176500	RS-03	8/23/2011	Normal	Replicate	RS-02	+	—	—	—	+	—	—	—		
04157000	SG-01	3/15/2011	Normal	Regular		+	—	—	—	—	—	—	—		
04157000	SG-05	6/16/2011	Normal	Regular		—	—	—	—	—	—	—	—		
04157000	SG-06	7/14/2011	Normal	Regular		—	—	—	—	—	—	—	—		
04157000	SG-07	7/14/2011	Normal	Replicate	SG-06	+	—	—	—	—	—	—	—		
04157000	SG-08	8/23/2011	Normal	Regular		+	—	—	—	—	—	—	—		
04157000	SG-11	9/14/2011	Normal	Regular		+	—	—	—	—	—	—	—		
04101500	SJ-02	4/7/2011	Normal	Regular		—	—	—	—	—	—	—	—		
04101500	SJ-07	7/27/2011	Normal	Regular		+	—	—	—	—	—	—	—		
04101500	SJ-08	8/24/2011	Normal	Regular		+ <sup>v</sup>	—	—	—	—	—	—	—		
04269000	SR-01	3/24/2011	Normal	Regular		—	—	+	—	—	—	—	—		
04269000	SR-04	4/26/2011	Normal	Regular		—	—	—	—	—	—	—	—		
04269000	SR-07	6/21/2011	Normal	Regular		+	—	—	—	—	—	—	—		
04269000	SR-08	7/26/2011	Normal	Regular		+	—	—	—	—	—	—	—		
04269000	SR-09	7/26/2011	Normal	Replicate	SR-08	+	—	—	—	—	—	—	—		
04269000	SR-13	9/20/2011	Normal	Regular		+	—	—	—	—	—	+	—		
04024000	ST-01	3/15/2011	Normal	Regular		—	—	—	—	—	—	—	—		
04024000	ST-02	4/20/2011	Normal	Regular		—	—	—	—	—	—	—	—		
04024000	ST-04	6/14/2011	Normal	Regular		+	+	—	—	—	—	—	—		
04024000	ST-05	6/23/2011	Normal	Regular		+	—	—	—	—	—	—	—		
04024000	ST-06	7/21/2011	Normal	Regular		—	—	—	—	—	—	—	—		
04024000	ST-07	7/21/2011	Normal	Replicate	ST-06	+	—	—	—	—	—	—	—		
04024000	ST-11	8/31/2011	Normal	Regular		—	—	—	—	—	—	—	+		
04024000	ST-12	9/13/2011	Normal	Regular		+	—	—	—	—	—	—	—		

**Appendix 3. Pathogen gene detection in Great Lakes tributaries, 2011.—Continued**

[Streamgaging stations corresponding to station IDs and field IDs are listed in table 1 in the main report. USGS, U.S. Geological Survey; Assoc. environ. sample, associated environmental sample; *E. coli*, *Escherichia coli*; +, gene present in sample; —, gene absent in sample; ns, no sample; V—remark code indicating possible contamination (analyte was detected in both the environmental sample and the associated blanks)]

USGS station ID	Field ID	Sample collection date	Flow conditions	Sample type	Assoc. environ. sample	E. coli gene data				Shigella gene data		Salmonella gene data		Campylobacter gene data	
						eaeA	stx2	stx1	rfb O157	ipaH	invA	spvC	<i>C. coli</i> / <i>C. jejuni</i>		
04137500	AU-02	4/12/2011	High	Regular		—	—	—	—	—	—	—	—	—	—
04027000	BD-01	4/11/2011	High	Regular		—	—	—	—	—	—	—	—	—	—
04027000	BD-02	4/11/2011	High	Replicate	BD-01	—	—	—	—	—	—	—	—	—	—
04027000	BD-03	5/24/2011	High	Regular		+	—	—	—	—	—	—	—	—	—
04095090	BN-02	4/28/2011	High	Regular		+	—	+	+	—	—	+	—	—	+
04095090	BN-03	5/25/2011	High	Regular		+	—	—	—	—	—	+	—	—	—
04095090	BN-04	6/8/2011	High	Regular		—	—	—	—	—	—	—	—	—	—
04095090	BN-10	9/7/2011	High	Regular		+	—	—	—	—	—	—	—	—	—
04165500	CL-01	5/16/2011	High	Regular		+	+	+	+	—	—	—	—	—	—
04165500	CL-02	8/17/2011	High	Regular		+	—	—	—	—	—	—	—	—	—
04165500	CL-03	8/17/2011	High	Replicate	CL-02	+	—	—	+	—	—	—	—	+	—
04208000	CY-01	4/27/2011	High	Regular		—	—	—	—	—	—	—	—	—	—
04208000	CY-02	7/26/2011	High	Regular		+	—	—	—	—	—	—	—	—	—
04208000	CY-03	7/26/2011	High	Replicate	CY-02	+	+	—	—	—	—	—	—	—	—
04208000	CY-07	9/20/2011	High	Regular		+	+	+	+	—	—	—	—	+	—
04208000	CY-08	10/26/2011	High	Regular		+	—	—	—	—	—	—	—	—	—
040851385	FX-01	5/4/2011	High	Regular		+	—	—	—	—	—	—	—	—	—
04119400	GR-01	4/29/2011	High	Regular		+	+	—	—	—	—	—	—	—	—
04119400	GR-02	5/18/2011	High	Regular		+	—	—	—	—	—	—	—	—	—
04119400	GR-03	6/2/2011	High	Regular		+	—	—	—	—	—	—	—	—	—
04119400	GR-04	6/29/2011	High	Regular		+	—	—	—	—	—	—	—	—	—
04119400	GR-05	6/29/2011	High	Replicate	GR-04	+	—	—	—	—	—	—	—	—	—
04119400	GR-06	8/2/2011	High	Regular		+	+	+	—	—	—	—	—	—	—
04231600	GS-02	4/26/2011	High	Regular		+	—	—	—	—	—	—	—	—	—
04231600	GS-03	5/17/2011	High	Regular		+	—	—	+	—	—	—	—	—	—

**Appendix 3.** Pathogen gene detection in Great Lakes tributaries, 2011.—Continued

[Streamgaging stations corresponding to station IDs and field IDs are listed in table 1 in the main report. USGS, U.S. Geological Survey; Assoc. environ. sample, associated environmental sample; *E. coli*, *Escherichia coli*; +, gene present in sample; —, gene absent in sample; ns, no sample; V—remark code indicating possible contamination (analyte was detected in both the environmental sample and the associated blanks)]

USGS station ID	Field ID	Sample collection date	Flow conditions	Sample type	Assoc. environ. sample	E. coli gene data					Shigella gene data		Salmonella gene data		Campylobacter gene data	
						eaeA	stx2	stx1	rfb O157	ipaH	invA	spvC	<i>C. coli</i> / <i>C. jejuni</i>			
04231600	GS-04	6/2/2011	High	Regular		+	+	—	—	—	—	—	—	—	—	
04193500	MA-01	3/8/2011	High	Regular		+	+	—	—	—	—	+	+	—	+	
04085427	MN-02	7/6/2011	High	Regular		+	—	—	—	—	—	—	—	—	—	
04087170	MW-01	5/3/2011	High	Regular		—	—	—	—	—	—	—	—	—	—	
04040000	ON-06	6/23/2011	High	Regular		+	—	—	+	—	—	—	—	—	—	
04249000	OS-01	3/9/2011	High	Regular		—	—	—	—	—	—	—	—	—	—	
04249000	OS-03	4/27/2011	High	Regular		—	+	—	—	—	—	—	—	—	—	
04249000	OS-04	5/9/2011	High	Regular		—	—	—	—	—	—	—	—	—	—	
04249000	OS-15	9/7/2011	High	Regular		+	—	—	—	—	—	—	—	—	—	
04195500	PO-01	3/8/2011	High	Regular		+	—	—	—	—	—	—	—	—	+	
04195500	PO-02	4/20/2011	High	Regular		+	+	—	+	—	—	+	—	—	—	
04195500	PO-03	7/27/2011	High	Regular		+	—	—	—	—	—	—	—	—	—	
04195500	PO-04	7/27/2011	High	Replicate	PO-03	+	—	—	—	—	—	—	—	—	—	
04102500	PP-02	4/21/2011	High	Regular		+	+	—	—	—	—	+	+	—	+	
04102500	PP-03	6/1/2011	High	Regular		+	+	—	—	—	—	ns	ns	—	—	
04102500	PP-04	7/19/2011	High	Regular		+	+	+	—	—	—	+	+	—	—	
04102500	PP-05	8/3/2011	High	Regular		+	+	—	—	—	—	—	—	—	—	
04102500	PP-08	10/4/2011	High	Regular		+	—	—	—	—	—	—	—	—	—	
04166500	RR-01	5/16/2011	High	Regular		+	+	+	—	—	—	—	+	—	—	
04166500	RR-02	5/26/2011	High	Regular		+	+	+	+	—	+	+	+	—	—	
04157000	SG-02	4/6/2011	High	Regular		+	—	—	+	—	—	—	—	—	—	
04157000	SG-03	5/2/2011	High	Regular		+	—	—	—	—	—	—	—	—	—	
04157000	SG-04	5/16/2011	High	Regular		+	+	—	—	—	—	—	—	—	—	
04101500	SJ-01	3/9/2011	High	Regular		+	—	—	—	—	—	—	—	—	+	
04101500	SJ-03	4/21/2011	High	Regular		+	+	+	+	—	—	+	—	—	—	





**Appendix 4.** Quality-control data—Pathogen gene detection in Great Lakes tributaries, 2011.—Continued

[Streamgaging stations corresponding to station IDs and field IDs are listed in table 1 in the main report. USGS, U.S. Geological Survey; Assoc. environ. sample, associated environmental sample; *E. coli*, *Escherichia coli*; +, gene present in sample; -, gene absent in sample; ns, no sample; v - value qualifier code (analyte detected in laboratory blank); V - value qualifier code (contamination, analyte was detected in both the environmental sample and the associated blanks)]

USGS station ID	Field ID	Sample collection Date	Sample type	Assoc. environ. sample	E. coli gene data				Shigella gene data	Salmonella gene data	Campylobacter gene data
					eaeA	stx2	stx1	rfb O157			
040851385	FX-05	9/19/2011	Source solution blank	ns	-	-	-	-	-	-	-
04119400	GR-08	10/31/2011	Source solution blank	ns	+	-	-	-	-	-	-
04231600	GS-12	7/18/2011	Source solution blank	GS-09	-	-	-	-	-	-	-
04193500	MA-06	7/27/2011	Source solution blank	MA-03	-	-	-	-	-	-	-
04067500	MM-06	9/19/2011	Source solution blank	ns	-	-	-	-	-	-	-
04085427	MN-06	9/19/2011	Source solution blank	ns	-	-	-	-	-	-	-
04087170	MW-05	9/20/2011	Source solution blank	MW-02	-	-	-	-	-	-	-
04040000	ON-11	9/20/2011	Source solution blank	ns	+	-	-	-	-	-	-
04249000	OS-13	7/18/2011	Source solution blank	OS-10	-	-	-	-	-	-	-
04195500	PO-06	7/27/2011	Source solution blank	PO-03	-	-	-	-	-	-	-
04166500	RR-06	10/31/2011	Source solution blank	ns	-	-	-	-	-	-	-
04176500	RS-05	8/30/2011	Source solution blank	ns	-	-	-	-	-	-	-
04157000	SG-10	8/23/2011	Source solution blank	SG-08	-	-	-	-	-	-	-
04101500	SJ-10	8/24/2011	Source solution blank	SJ-08	+	-	-	-	-	-	-
04269000	SR-11	7/26/2011	Source solution blank	SR-08	-	-	-	-	-	-	-
04024000	ST-09	7/21/2011	Source solution blank	ST-06	-	-	-	-	-	-	-



